

Available online at www.sciencedirect.com



PHARMACOLOGY BIOCHEMISTRY <sup>AND</sup> BEHAVIOR

Pharmacology, Biochemistry and Behavior 74 (2003) 869-876

www.elsevier.com/locate/pharmbiochembeh

# Involvement of the sigma<sub>1</sub> receptor in the motivational effects of ethanol in mice

Tangui Maurice\*, Magali Casalino, Magali Lacroix, Pascal Romieu<sup>1</sup>

Behavioral Neuropharmacology Group, Institut de Biologie, INSERM U. 336, 4, bvd Henri IV, 34060 Montpellier, France

Received 9 September 2002; received in revised form 17 December 2002; accepted 17 December 2002

#### Abstract

In the present study, we examined the involvement of the sigma<sub>1</sub> ( $\sigma_1$ ) receptor in several behavioral manifestations of ethanol addiction. Administration of ethanol (0.5, 1, and 2 g/kg) in Swiss mice dose-dependently induced locomotor stimulation, conditioned place preference, and conditioned taste aversion, which are considered as behavioral index of drug-induced reward. Pretreatment with the selective  $\sigma_1$  receptor antagonist *N*-[2-(3,4-dichlorophenyl)ethyl]-*N*-methyl-2-(dimethylamino)ethylamine (BD1047, 3–30 mg/kg) dose-dependently blocked ethanol (1 g/kg)-induced hyperlocomotion and taste aversion and ethanol (2 g/kg)-induced place preference. Pretreatment with the selective  $\sigma_1$  receptor agonist 2-(4-morpholino)ethyl 1-phenylcyclohexane-1-carboxylate (PRE-084, 1–10 mg/kg) before ethanol (0.5 g/kg) failed to affect the resulting locomotor stimulation, but dose-dependently enhanced the conditioned place preference. Each  $\sigma_1$  receptor ligand was devoid of behavioral effect by itself. These observations show that activation of the  $\sigma_1$  receptor is a necessary component of ethanol-induced motivational effects and suggest a new pharmacological target for alleviating ethanol addiction. © 2003 Elsevier Science Inc. All rights reserved.

*Keywords:* Ethanol; Sigma<sub>1</sub> ( $\sigma_1$ ) receptor; Hyperlocomotion; Conditioned place preference; Reward; Conditioned taste aversion; Mouse

# 1. Introduction

Activation and plasticity of the mesolimbic dopaminergic system contribute, as a common neurobiological process, to the acquisition and expression of the rewarding effects of drugs of abuse (Di Chiara, 1998). This has been extensively detailed for dopaminergic abused drugs, including amphetamine or cocaine (Kuhar, 1992; Koob, 1996). It is also true for opiate drugs like morphine, cannabinoids, and ethanol. Although several neurotransmitter systems contribute to ethanol reward and dependence, dopaminergic pathways are prominent substrates for the neuroadaptations involved (Tabakoff and Hoffman, 1996; Koob et al., 1998). Indeed, systemic administration or oral self-administration of ethanol stimulated dopamine release preferentially in the nucleus accumbens and bed nucleus of stria terminalis of rats (Di Chiara and Imperato, 1988; Weiss et al., 1993; Carboni et al., 2000). Moreover, ethanol reward or self-administration

could be modulated using dopamine receptor agonists or antagonists (Samson et al., 1993a,b; Boyce and Risinger, 2000). In addition, dopamine D2 receptor knock-out mice failed to develop ethanol-induced conditioned place preference, confirming that dopamine D2 receptors are involved in ethanol reward (Cunningham et al., 2000). Therefore, pharmacological strategies may consist in treatments with drugs acting directly on dopaminergic receptors, which include D1-like receptors, namely D1 and D5, and D2-like receptors, namely D2, D3, and D4 (Civelli et al., 1993). Both D1- and D2-like receptors are present in reward-related brain structures, including the olfactory bulb, ventral tegmental area, nucleus accumbens, amygdala, frontal cortex, septum, and bed nucleus of stria terminalis (Koob, 1992; Civelli et al., 1993; Carboni et al., 2000). In turn, effective direct strategies may use dopaminergic antagonists showing a very high selectivity in vivo, and this has been still difficult to achieve. Alternate strategies could also be proposed through effective neuromodulatory systems. In particular, the sigma<sub>1</sub>  $(\sigma_1)$  receptor may represent such putative target.

The  $\sigma_1$  receptor, localized intracellularly within neurons, is a 223-amino acid protein, cloned in several animal species and humans (Hanner et al., 1996; Kekuda et al., 1996; Seth et al., 1997, 1998; Pan et al., 1998). The  $\sigma_1$  receptor appeared

<sup>\*</sup> Corresponding author. CNRS UMR 5102, Université de Montpellier II, cc 090, place Eugène Bataillon, 34095 Montpellier Cedex, France. Tel.: +33-4-67-14-42-70; fax: +33-4-67-14-42-51.

E-mail address: maurice@univ-montp2.fr (T. Maurice).

<sup>&</sup>lt;sup>1</sup> Present address: CNRS UMR 5102, Université de Montpellier II, cc 090, place Eugène Bataillon, 34095 Montpellier Cedex, France.

devoid of analogy with any other known mammalian protein. Its regional distribution, determined in rodents using in situ hybridization or immunohistochemistry (Zamanillo et al., 2000; Kitaichi et al., 2000; Alonso et al., 2000), showed moderate to intense staining in most of the dopaminergic structures, including the caudate-putamen, nucleus accumbens, amygdala, septum, and frontal cortex (Alonso et al., 2000; Maurice et al., 2002). The  $\sigma_1$  receptor mediates a potent modulation of several neurotransmitter systems by affecting intracellular second messengers systems, particularly Ca<sup>2+</sup> mobilization (Hayashi et al., 2000; Hayashi and Su, 2001). Noteworthy,  $\sigma_1$  ligands efficiently modulate the dopaminergic neurotransmission and several studies reported the effect of  $\sigma_1$  ligands on dopamine synthesis, metabolism, and release or electric activity of dopaminergic neurons (for review, see Maurice et al., 2002).

The  $\sigma_1$  receptor has recently been demonstrated to be involved in cocaine's rewarding effects (Romieu et al., 2000, 2002). The  $\sigma_1$  receptor is involved in the acquisition or expression of cocaine-induced conditioned place preference in mice, as demonstrated using selective  $\sigma_1$  receptor antagonists or antisense oligodeoxynucleotide probes. We also observed that a repeated 4-day cocaine treatment increased the expression of the  $\sigma_1$  receptor mRNA in the nucleus accumbens, suggesting that the drug is able to induce overexpression of  $\sigma_1$  gene and that neuroadaptations involving the  $\sigma_1$  receptor occurred in mesolimbic structures involved in addiction (Romieu et al., 2002).

In the present study, we characterized the involvement of the  $\sigma_1$  receptor in the motivational effects of ethanol in Swiss mice. We used different behavioral procedures including locomotor activity measures, conditioned place preference, and taste aversion. Place conditioning is frequently used to index the rewarding properties of abused drugs (Carr et al., 1989; Tzschentke, 1998; Romieu et al., 2000, 2002). Conditioned taste aversion is produced by most of abused drugs. It has been proposed to be related with the sensitivity to drug reward (Hunt and Amit, 1987). Animals were thus treated with ethanol and/or the selective  $\sigma_1$  receptor antagonist N-[2-(3,4-dichlorophenyl)ethyl]-N-methyl-2-(dimethylamino)ethylamine (BD1047) (Matsumoto et al., 1995) or the selective  $\sigma_1$  receptor agonist 2-(4-morpholino)ethyl 1phenylcyclohexane-1-carboxylate (PRE-084) (Su et al., 1991). We investigated here whether treatment with selective  $\sigma_1$  antagonist or agonist affect the ethanol-induced effects on all behavioral measures in order to examine whether the  $\sigma_1$  receptor activation is involved in acquisition of the motivational effects and reward induced by ethanol.

# 2. Methods

#### 2.1. Subjects

Male Swiss OF1 mice (Breeding Center of the Faculty of Pharmacy, Montpellier, France), aged 5–6 weeks and weigh-

ing  $30\pm 2$  g, were used. Animals were housed in plastic cages in groups. They had free access to laboratory food and water, except during behavioral experiments, and they were kept in a regulated environment ( $23\pm 1$  °C, 40-60% humidity) under a 12-h light–dark cycle (light on at 7:00 a.m.). Animals submitted to taste conditioning had their access to fluids restricted as described herein. Experiments were carried out between 9:00 a.m. and 5:00 p.m. in a sound-attenuated and air-regulated experimental room to which mice were habituated at least 30 min before each experiment. All animal procedures were conducted in strict adherence with the European Community Council Directive of 24 November 1986 (86-609/EEC).

### 2.2. Drugs

Ethanol (Carlo Erba, Rodano, Italy) was diluted in saline solution to the final concentrations (200 and 400 g/l corresponding to the 1- and 2-g/kg doses, respectively) by taking into account the density (0.79). BD1047 was given by Dr. Wayne D. Bowen (NIDDK, NIH, Bethesda, MD, USA) and PRE-084 by Dr. Tsung-Ping Su (IRP, NIDA, NIH, Baltimore, MD, USA). BD1047 and PRE-084 were diluted in saline solution. Injections were performed through the intraperitoneal route in a volume of 100  $\mu$ l/20 g body weight.

#### 2.3. Locomotor activity

Mice were placed in a Plexiglas cage  $(25 \times 40 \times 15 \text{ cm})$  high) for 40 min, between t=0 and t=40 min. The  $\sigma_1$  receptor ligand, BD1047 or PRE-084, was injected at t=30 and ethanol or saline was injected at t=40 min. Animals were again placed in the cage for 40 min, between t=40 and t=80 min. The locomotion was recorded using infrared light sources and detectors positioned opposite to each other at 1-in. intervals on the walls of the monitoring system (Opto-Varimex, Columbus Instruments, Columbus, OH, USA). Locomotion was measured in terms of infrared light beams interrupted over 10-min periods, between t=40 and t=80 min (time-course profiles), and summed over 30 min, between t=50 and t=80 min (global activity scores).

#### 2.4. Conditioned place preference

The apparatus consisted of a PVC box divided into two compartments of equal size  $(15 \times 15 \times 35 \text{ cm high})$  separated by sliding doors. The first compartment had black walls and floor, the second one had white walls and floor. Each compartment presented different floor textures, smooth for the black one and covered by a wire mesh grid for the white. A 60-W lamp lit the white compartment during all experiments.

The procedure consisted of three different phases (Romieu et al., 2000, 2002): preconditioning (Day 1), conditioning (Days 2-5), and postconditioning (Day 6). For the preconditioning phase, each mouse was placed in the white

compartment and after 5 s, the sliding doors were raised. The animal was allowed to freely explore the apparatus for 10 min. The preconditioning phase was repeated after 6 h. The exploration was videotaped and the amount of time spent in each compartment was determined in order to assess the unconditioned preference. Animals showing a strong unconditioned preference (>570 s), i.e., less than 5%, were discarded. Place preference conditioning was conducted using an unbiased procedure. In each experimental group, half of the animals received ethanol in the spontaneously preferred compartment and the other half in the nonpreferred compartment. Immediately after the drug injection, each mouse was confined to the drug-paired compartment for 15 min. In coadministration experiments, mice received the  $\sigma_1$  receptor ligand 10 min before ethanol. After a 6-h washout period, they were administered the vehicle solutions and confined to the other compartment for 15 min. The postconditioning test was performed the last day. Each mouse was again placed in the white compartment and after 5 s, the doors were raised. The animal was allowed to freely explore the apparatus for 10 min. The exploration was videotaped and the amount of time spent in each compartment was determined. The conditioned score represented the difference of time spent in the drug-paired compartment between the post- and preconditioning sessions.

#### 2.5. Conditioned taste aversion

The taste conditioning test was conducted in home cages, animals being housed by groups of five per cage (n = 10 cages per treatment group). Fluids were presented at room temperature in 50-ml graduated glass cylinders fitted with stainlesssteel drinking spouts inserted through the grid of the cage. Consumption was measured over 1-h periods by weighing the drinking tubes and was corrected for evaporation and spillage by subtracting the mean fluid loss measured in three drinking tubes placed onto an empty cage for an equal amount of time. Subjects were adapted to a water restriction regimen over a 7-day period, with 2-h of water per day from 9:00 to 11:00 a.m. At 48-h intervals over the next 10 days, mice had access to a 0.2-M NaCl solution between 9:00 and 10:00 a.m. (Risinger and Cunningham, 1992; Risinger et al., 1999, 2001). For the first four trials, immediately after access to the NaCl solution, each mouse was administered intraperitoneally with BD1047 (0, 3, 10, or 30 mg/kg) and ethanol (0 or 1 g/kg). Animals also received 30-min access to tap water 5 h after each NaCl access period, to prevent dehydration. On the final trial, animals were given access to the NaCl solution without further injection. On intervening days, mice had a 2-h access to water (9:00-11:00 a.m.). Experimental data represented the mean NaCl solution intake per cage.

#### 2.6. Statistical analyses

All measures (locomotor activity counts, conditioned scores, or NaCl solution intakes) were expressed as mean  $\pm$ 

S.E.M. and analyzed using the Newman–Keuls or Dunnett tests for multiple comparisons after parametric one- or twoway analyses of variance (ANOVA, F values). The criterion for statistical significance was P < .05.

#### 3. Results

## 3.1. Locomotor activity

Intraperitoneal injections of ethanol induced in Swiss mice a dose-dependent stimulation of locomotor activity, as measured using infrared beam interruptions. Locomotion was measured every 10 min after injections for 40 min and global activity was determined as the total activity during the last 30 min. Vehicle-treated animals showed a global activity in the 741- to 755-count range (n=12-13, Figs. 1A and 2A) and activity decreased regularly with time [ANOVA: F(3,47) =6.63, P < .01, and test for linear trend: F = 19.28, P < .0001, open circles, Fig. 1B; ANOVA: F(3,51) = 5.66, P < .01, and test for linear trend: F = 16.47, P < .001, open circles, Fig. 2B]. The ethanol treatment increased this score to  $1289 \pm 89$ (n = 14, P < .05 vs. vehicle-treated group, Fig. 1A) at the dose of 0.5 g/kg and  $1794 \pm 89$  (n = 12, P < .01 vs. vehicle-treated group, Fig. 2A) at the dose of 1 g/kg. This increase corresponded to a similar augmentation of the activity over the 40 min of measure and activity decreased regularly with time [ANOVA: F(3,55) = 32.96, P < .0001, and test for linear trend: F = 7.05, P < .0001, closed circles, Fig. 2B].

The effect of the selective  $\sigma_1$  receptor agonist PRE-084 was first examined, in the 1- to 10-mg/kg dose range, on the locomotor increase induced by 0.5 g/kg ethanol. The drug failed to affect the locomotor response when administered alone (Fig. 1A). The hyperlocomotion was observed in all groups treated with ethanol [F(7,103) = 2.54, P < .01], and the PRE-084 pretreatment failed to induce any significant change (Fig. 1A,B). The effect of the selective  $\sigma_1$  receptor antagonist BD1047 was then examined in the 3- to 30-mg/kg dose range on the locomotor increase induced by 1 g/kg ethanol. The drug failed to affect the locomotor response when administered alone (Fig. 2A). However, the hyperlocomotion observed in the ethanol-treated group was diminished in a dose-dependent manner by the BD1047 pretreatment [F(7,95) = 5.44, P < .0001]. In particular, at the highest dose tested, BD1047 allowed a complete blockade of the ethanol (1 g/kg)-induced locomotor increase, in terms of global activity (Fig. 2A) or time-course profile (Fig. 2B).

#### 3.2. Conditioned place preference

Repeated administration of ethanol (0.5–2 g/kg) over 4 days in Swiss mice and confinement in the drug-paired compartment led to the development of a dose-dependent place preference. Indeed, a significantly positive conditioned score was measured [F(3,76)=3.63, P<.05, Fig. 3A].



Fig. 1. Effect of the selective  $\sigma_1$  receptor agonist PRE-084 on ethanol-induced locomotor hyperactivity: (A) total activity during the last 30 min of the session and (B) time-course. Each mouse was placed in the experimental cage for 40 min. PRE-084 (1, 3, and 10 mg/kg ip) or the vehicle solution (Veh) was injected 10 min before ethanol (0.5 g/kg ip). Locomotor activity was measured immediately after ethanol injection in terms of number of occlusions of infrared light beams every 10 min during 40 min. The number of animals per group is indicated within the columns in (A). \*P < .05, \*\*P < .01 vs. vehicle-treated group, Newman–Keuls' test.

Previous studies, using the same experimental procedure, showed that the selective  $\sigma_1$  receptor ligands, PRE-084 and BD1047, failed to induce conditioned place preference when injected alone (Romieu et al., 2000, 2002). However, pre-administration of PRE-084 in the 1- to 3-mg/kg dose range dose-dependently facilitated the development of place preference after treatment with a low, subactive dose of 0.5 g/kg

ethanol [F(2,52)=3.74, P<.05, Fig. 3B]. In particular, animals treated with 3 mg/kg PRE-084 and ethanol acquired place preference, the conditioned score being significantly higher than mice treated with ethanol acquired place preference, the conditioned score being significantly higher than mice treated with ethanol alone (Fig. 3B). Moreover, pre-administration of BD1047 in the 3- to 10-mg/kg dose range



Fig. 2. Effect of the selective  $\sigma_1$  receptor antagonist BD1047 on ethanol-induced locomotor hyperactivity: (A) total activity during the last 30 min of the session and (B) time-course. Each mouse was placed in the experimental cage for 40 min. BD1047 (3, 10, and 30 mg/kg ip) or the vehicle solution (Veh) was injected 10 min before ethanol (1 g/kg ip). Locomotor activity was measured immediately after ethanol injection in terms of number of occlusions of infrared light beams every 10 min during 40 min. The number of animals per group is indicated within the columns in (A). \**P*<.05, \*\**P*<.01 vs. the vehicle-treated group; ##*P*<.01 vs. the ethanol-treated group, Newman–Keuls' test.



Fig. 3. Effect of selective  $\sigma_1$  receptor ligands on acquisition of ethanol-induced conditioned place preference: (A) dose-response effect of ethanol, (B) pretreatment of the selective  $\sigma_1$  receptor agonist PRE-084, and (C) pretreatment of the selective  $\sigma_1$  receptor antagonist BD1047. The  $\sigma_1$  receptor ligands were administered intraperitoneally 10 min before ethanol (0–2 g/kg ip), which was given immediately before placement in the compartment during the conditioning test. The conditioned score represents the difference in time spent in the drug-paired compartment between the post- and preconditioning sessions. The number of animals per group is indicated between parentheses. \**P*<.01 vs. the vehicle (Veh)-treated group, <sup>#</sup>*P*<.05 vs. the ethanol-treated group, Dunnett's test.

dose-dependently inhibited the acquisition of place preference induced by 2 g/kg ethanol [F(2,55) = 6.13, P < .01, Fig. 3C]. Animals treated with 10 mg/kg BD1047 and ethanol failed to show any place preference (Fig. 3C). In other words, pretreatment with the  $\sigma_1$  receptor agonist facilitated acquisition of ethanol-induced reward, while pretreatment with the  $\sigma_1$  antagonist blocked ethanol-induced reward.



Fig. 4. Effect of the selective  $\sigma_1$  receptor antagonist BD1047 on ethanol-induced conditioned taste aversion. Mean 0.2 M NaCl solution intake were measured during 1 h. On Days 1, 3, 5, 7, and 9, after the 1-h access to NaCl solution, groups received either saline solution (Veh) or BD1047(3 and 10 mg/kg ip) and/or ethanol (1 g/kg ip). Animals received 2-h access to tap water on intervening days. The number of animals per group was n = 10. \*P < .05, \*\*P < .01 vs. the vehicle-treated group on the same day,  ${}^{\#}P < .05$ ,  ${}^{\#}P < .01$  vs. the same group on Day 1, Newman–Keuls' test.

#### 3.3. Conditioned taste aversion

Ethanol (1 g/kg) injection gradually reduced NaCl solution intake on Days 3, 5, 7, and 9, indicating the development of conditioned taste aversion (Fig. 4). This reduction was not due to a decrease in liquid intake because a preliminary experiment showed that replacing NaCl solution by tap water resulted in a similar liquid intake by vehicle- or ethanol-treated animals (data not shown). The  $\sigma_1$  receptor antagonist BD1047 failed to affect NaCl solution intake by itself. However, pretreatment with BD1047 dose-dependently blocked the ethanol-induced decrease in NaCl solution intake. Treatment × Day analysis showed significant effects for treatment [F(7,360) = 119.79, P < .001], day [F(4,360) = 32.80, P < .001].001], and Treatment × Day [F(28,360) = 11.42, P < .001]. As shown in Fig. 4, NaCl intake measured for the group treated with BD1047 (30 mg/kg) and ethanol were similar as those measured for the vehicle-treated group on all days.

# 4. Discussion

The present pharmacological study showed that selective  $\sigma_1$  receptor ligands potently modulated the motivational effects of ethanol in the Swiss mouse. First, the  $\sigma_1$  antagonist BD1047 dose-dependently attenuated ethanol-induced locomotor activity. Second, the  $\sigma_1$  agonist PRE-084 facilitated, while BD1047 blocked, the acquisition of ethanol-induced conditioned place preference. Third, BD1047 blocked the acquisition of ethanol-induced conditioned taste aversion. The  $\sigma_1$  receptor is a unique intraneuronal protein, mediating an effective and wide-range neuromodulatory action in the brain. In particular, a recent hypothesis suggests that it acts a sensor/modulator of intracellular Ca<sup>2+</sup> mobilization and homeostasis on the endoplasmic reticulum membrane because activation of the receptor by agonists, including synthetic compounds, such as PRE-084 or cocaine, or endogenous steroids, such as dehydroepiandrosterone or pregnenolone sulfates, enhanced binding of inositol 1,4,5-triphosphate (InsP<sub>3</sub>) to its receptor and  $Ca^{2+}$  mobilization from InsP<sub>3</sub> receptor-sensitive pools (Hayashi et al., 2000; Hayashi and Su, 2001; Su and Hayashi, 2001). Consequently, the  $\sigma_1$  receptor activation has been implicated in several neuroadaptative changes, as observed in learning and memory, depression, schizophrenia, and reward and behavioral sensitization induced by cocaine or methamphetamine (Maurice et al., 1999, 2001, 2000). Indeed, cocaine- and methamphetamineinduced hyperlocomotor activity, stereotyped behaviors, and sensitization of these behavioral responses after repeated administration could be blocked by the selective  $\sigma_1$  receptor antagonists  $\alpha$ -(4-fluorophenyl)-4-(5-fluoro-2-pyrimidinyl)-1-piperazine butanol (BMY-14,802), rimcazole, 6-[6-(4-hydroxypiperidinyl)hexyloxy]-3-methylflavone (NPC 16377), BD1008, BD1047, or 1-[2-(3,4-dichloro-phenyl)ethyl]-4methylpiperazine (BD1063) (Menkel et al., 1991; Witkin et al., 1993; Ujike et al., 1992, 1996; McCracken et al., 1999). More recently, activation of the  $\sigma_1$  receptor was also demonstrated in the acquisition and expression of cocaine-induced conditioned place preference in C57BL/6 mice (Romieu et al., 2000, 2002). Activation of the  $\sigma_1$  receptor may thus result from the activation of dopaminergic systems, particularly in mesolimbic structures, and may, conversely, mediate the modulation of dopaminergic tonus (for a review, see Maurice et al., 2002).

In mice, ethanol produces locomotor stimulation at doses in the 0.5 to 2 g/kg ip range (Risinger and Oakes, 1996). A positive correlation between ethanol-induced locomotor stimulation and ethanol-induced reward has been evoked through the euphorigenic effects of ethanol as part of the rewarding properties (Wise and Bozart, 1987; Phillips and Shen, 1996). The dopaminergic involvement in ethanolinduced locomotor stimulation was suggested by studies showing that the D2 dopamine receptor antagonist haloperidol blocked the response (Risinger et al., 1992). It must be outlined that haloperidol is also the most potent  $\sigma_1$  receptor antagonist known to date, and the putative involvement of the  $\sigma_1$  receptor blocked in such effect is unresolved. We observed here that BD1047 blocked the ethanol-induced hyperlocomotor activity, indicating that the  $\sigma_1$  receptor is involved in this acute response to ethanol. Activation of the mesolimbic dopaminergic pathways, particularly within the nucleus accumbens has been evoked as responsible of this behavioral response, and  $\sigma_1$  receptors are highly present within this structure (Maurice et al., 2002). In addition, we previously observed that a 4-day treatment of C57BL/6 mice with cocaine (20 mg/kg) resulted in an increased expression of the  $\sigma_1$  receptor selectively in the nucleus accumbens (Romieu et al., 2002), suggesting that  $\sigma_1$  receptors located within this key structure are particularly sensitive to psychostimulants.

Ethanol produces conditioned place preference (Cunningham et al., 1992, 2000; Risinger and Oakes, 1996), and this behavioral procedure is routinely considered as a reliable index of neurophysiological mechanisms involved in drug reward (Schechter and Calcagnetti, 1993). A crucial role for the  $\sigma_1$  receptor activation was clearly observed here because the  $\sigma_1$  agonist was able to potentiate acquisition of place preference while the  $\sigma_1$  antagonist blocked it. The pharmacological systems sustaining ethanol-induced place preference are still under question. Serotogenic and GABAergic systems, rather than dopaminergic or opioid systems, have been involved in the acquisition of ethanol-induced place preference (Risinger et al., 2001). Pharmacological studies must be performed in order to determine whether an effect of  $\sigma_1$ receptors present within mesolimbic dopaminergic neurons can be ruled out for acquisition of place preference by ethanol, contrarily to what has been evoked for cocaine (Romieu et al., 2000, 2002).

Conditioned taste aversion is a particularly interesting behavioral response because it reflects a highly integrated response observed in alcoholics. Pairing a distinctive flavor with exposure to ethanol leads to the development of a conditioned aversion for the flavor (Hunt and Amit, 1987;

Sherman et al., 1998). Pharmacological studies implicated dopaminergic systems in ethanol-induced conditioned taste aversion through mainly D2, but also D1 and D4 receptors (Risinger et al., 1999, 2001). In the present study, ethanol provoked conditioned taste aversion, which is observable after the ethanol administration as soon as the second day of measure. BD1047 blocked acquisition of ethanol-induced conditioned taste aversion, in line with previous observations. The present study thus showed a similar involvement of the  $\sigma_1$  receptor in three different behavioral responses to ethanol. In particular, we did not observe a pharmacological dissociation between ethanol-stimulated activity and acquisition of conditioned place preference as evoked by Risinger et al. (2001). The wide-range neuromodulatory effect induced by  $\sigma_1$  receptor ligands suggested that this receptor affects several kind of neurotransmission systems, including glutamate-, acetylcholine-, dopamine-, serotonin-, and noradrenaline-induced responses (Maurice et al., 1999, 2001). At the physiological level, the  $\sigma_1$  receptor may be involved in the modulation of particular neuroadaptative changes, such as behavioral sensitization or reward, rather than particular neurochemical pathways. Consequently, it may be involved in the common neuroadaptive processes induced by cocaine or ethanol.

The precise brain structures involved in the  $\sigma_1$  receptormediated effect, and particularly the participation of mesolimbic dopaminergic systems, must however be examined. The mechanism of the  $\sigma_1$  receptor activation within these structures must be determined because at present there are no data suggesting a direct effect of ethanol on the  $\sigma_1$ receptor. The role of the  $\sigma_1$  receptor in ethanol tolerance, dependence, and withdrawal must also be examined. The  $\sigma_1$ receptor has been proposed to play a particular role in neuronal plasticity and long-term changes (Maurice et al., 1999; Su and Hayashi, 2001). Prolonged or repeated ethanol intake leads to reduced behavioral response to ethanol, i.e., to cellular tolerance. Ethanol withdrawal is characterized by the development of anxiety, depressive responses, and increased seizures susceptibility. The  $\sigma_1$  receptor may be putatively involved in the resulting changes in expression and function appearing in these neuroadaptive responses as soon as acute or repeated injections.

In summary, this study brought evidence that activation of the  $\sigma_1$  receptor is a necessary component for the acquisition of ethanol-induced reward and related behavioral manifestations. A selective antagonist, BD1047, allowed to block ethanol-induced locomotor stimulation, conditioned place preference, and conditioned taste aversion. Acting through the  $\sigma_1$  receptor may offer a new pharmacological strategy to counteract ethanol addiction.

#### Acknowledgements

The authors wish to thank Drs. A.J. Roberts and R. Martin-Fardon (Scripps Research Institute, La Jolla, CA, USA) for helpful advice, Dr. F.J. Roman (Pfizer GRD, Fresnes) for his appreciated help, and Drs. W.D. Bowen and T.-P. Su for their gift of drugs. This work was supported by INSERM.

#### References

- Alonso G, Phan VL, Guillemain I, Saunier M, Legrand A, Anoal M, et al. Immunocytochemical localization of the  $\sigma_1$  receptor in the adult rat central nervous system. Neuroscience 2000;97:155–70.
- Boyce JM, Risinger FO. Enhancement of ethanol reward by dopamine D3 receptor blockade. Brain Res 2000;880:202–6.
- Carboni E, Silvagni A, Rolando MTP, Di Chiara G. Stimulation of in vivo dopamine transmission in the bed nucleus of stria terminalis by reinforcing drugs. J Neurosci 2000;20(1-5):RC102.
- Carr GD, Fibiger HC, Philips AG. Conditioned place preference as a measure of drug reward. In: Liebman JM, Cooper SJ, editors. Neuropharmacological basis of reward. New York: Oxford Univ. Press; 1989. p. 264–319.
- Civelli O, Bunzow JR, Grandy DK. Molecular diversity of the dopamine receptors. Annu Rev Pharmacol Toxicol 1993;32:281–307.
- Cunningham CL, Neihus DR, Malott DH, Prather LK. Genetic differences in the rewarding and activating effects of morphine and ethanol. Psychopharmacology 1992;120:28–41.
- Cunningham CL, Howard MA, Gill SJ, Rubinstein M, Low MJ, Grandy DK. Ethanol-conditioned place preference is reduced in dopamine D2 receptor-deficient mice. Pharmacol Biochem Behav 2000;67:693–9.
- Di Chiara G. A motivational learning hypothesis of the role of mesolimbic dopamine in compulsive drug use. J Psychopharmacol 1998;12: 54–67.
- Di Chiara G, Imperato A. Drugs abused by humans preferentially increase synaptic dopamine concentrations in the mesolimbic system of freely moving rats. Proc Natl Acad Sci USA 1988;85:5274–8.
- Hanner M, Moebius FF, Flandorfer A, Knaus HG, Striessnig J, Kempner E, et al. Purification, molecular cloning, and expression of the mammalian sigma<sub>1</sub>-binding site. Proc Natl Acad Sci USA 1996;93:8072–7.
- Hayashi T, Su TP. Regulating ankyrin dynamics: roles of sigma-1 receptors. Proc Natl Acad Sci USA 2001;98:491–6.
- Hayashi T, Maurice T, Su TP. Ca<sup>2+</sup> signaling via sigma<sub>1</sub>-receptors: novel regulatory mechanism affecting intracellular Ca<sup>2+</sup> concentration. J Pharmacol Exp Ther 2000;293:788–98.
- Hunt T, Amit Z. Conditioned taste aversion induced by self-administered drugs: paradox revisited. Neurosci Biobehav 1987;11:107–30.
- Kekuda R, Prasad PD, Fei YJ, Leibach FH, Ganapathy V. Cloning and functional expression of the human type 1 sigma receptor (hSigmaR1). Biochem Biophys Res Commun 1996;229:553–8.
- Kitaichi K, Chabot JG, Moebius FF, Flandorfer A, Glossman H, Quirion R. Expression of the purported sigma<sub>1</sub> ( $\sigma_1$ ) receptor in the mammalian brain and its possible relevance in deficits induced by antagonism of the NMDA receptor complex as revealed using and antisense strategy. J Chem Neuroanat 2000;20:375–87.
- Koob GF. Drug of abuse: anatomy, pharmacology and function of reward pathways. Trends Pharmacol Sci 1992;13:177–84.
- Koob GF. Drug addiction: the yin and yang of hedonic homeostasis. Neuron 1996;16:893-6.
- Koob GF, Roberts AJ, Schulteis G, Parsons LH, Heyser CJ, Hyytia P, et al. Neurocircuitry targets in ethanol reward and dependence. Alcohol Clin Exp Res 1998;22:3–9.
- Kuhar MJ. Molecular pharmacology of cocaine: a dopamine hypothesis and its implications. Ciba Found Symp 1992;166:81–9.
- Matsumoto RR, Bowen WD, Tom MA, Vo VN, Truong DD, De Costa BR. Characterization of two novel sigma receptor ligands: antidystonic effects in rats suggest sigma receptor antagonism. Eur J Pharmacol 1995; 280:301–10.
- Maurice T, Phan VL, Urani A, Kamei H, Noda Y, Nabeshima T. Neuroactive neurosteroids as endogenous effector for the sigma<sub>1</sub> ( $\sigma_1$ ) receptor:

pharmacological evidences and therapeutic opportunities. Jpn J Pharmacol 1999;81:125-55.

- Maurice T, Urani A, Phan VL, Romieu P. The interaction between neuroactive steroids and the sigma<sub>1</sub> ( $\sigma_1$ ) receptor function: behavioral consequences and therapeutic opportunities. Brain Res Rev 2001;37:116–32.
- Maurice T, Martin-Fardon R, Romieu P, Matsumoto RR. Selective sigma<sub>1</sub> ( $\sigma_1$ ) receptor antagonists as a new promising strategy against cocaine addiction. Neurosci Biobehav Res 2000;26:499–527.
- McCracken KA, Bowen WD, Matsumoto RR. Novel  $\sigma$  receptor ligands attenuate the locomotor stimulatory effects of cocaine. Eur J Pharmacol 1999;365:35–8.
- Menkel M, Terry M, Ponteorvo M, Katz JL, Witkin JM. Selective  $\sigma$  ligands block stimulant effects of cocaine. Eur J Pharmacol 1991;201:251–2.
- Pan YX, Mei J, Xu J, Wan BL, Zuckerman A, Pasternak GW. Cloning and characterization of a mouse σ<sub>1</sub> receptor. J Neurochem 1998;70:2279–85.
- Phillips TJ, Shen EH. Neurochemical bases of locomotion and ethanol stimulant effects. Int Rev Neurobiol 1996;39:243–82.
- Risinger FO, Cunningham CL. Genetic differences in ethanol-induced hyperglycemia and conditioned taste aversion. Life Sci 1992;50:113-8.
- Risinger FO, Oakes RA. Dose- and conditioning trial-dependent ethanolinduced conditioned place preference in Swiss–Webster mice. Pharmacol Biochem Behav 1996;55:117–23.
- Risinger FO, Dickinson SD, Cunningham CL. Haloperidol reduces ethanolinduced motor activity stimulation but not conditioned place preference. Psychopharmacology 1992;107:453–6.
- Risinger FO, Brown MM, Oakes RA, Love JA. Effect of haloperidol or SCH-23390 on ethanol-induced conditioned taste aversion. Alcohol 1999;18:139–45.
- Risinger FO, Freeman PA, Greengard P, Fienberg AA. Motivational effects of ethanol in DARPP-32 knock-out mice. J Neurosci 2001;21:340–8.
- Romieu P, Martin-Fardon R, Maurice T. Involvement of the  $\sigma_1$  receptor in the cocaine-induced conditioned place preference. NeuroReport 2000;11: 2885–8.
- Romieu P, Phan VL, Martin-Fardon R, Maurice T. Involvement of the sigma<sub>1</sub> receptor in cocaine-induced conditioned place preference: possible dependence on dopamine uptake blockade. Neuropsychopharmacology 2002;26:444–55.
- Samson HH, Hodge CW, Tolliver GA, Haraguchi M. Effect of dopamine agonists and antagonists on ethanol-reinforced behavior: the involvement of the nucleus accumbens. Brain Res Bull 1993a;30:133–41.
- Samson HH, Tolliver GA, Pfeffer AD, Sadeghi K, Haraguchi M. Relation of ethanol self administration to feeding and drinking in a nonrestricted access situation in rats initiated to self-administer ethanol using the sucrose-fading technique. Alcohol 1993b;5:375–85.

- Schechter MD, Calcagnetti DJ. Trends in place preference conditioning with a cross indexed bibliography; 1975–1991. Neurosci Biobehav Rev 1993;17:21–41.
- Seth P, Leibach FH, Ganapathy V. Cloning and structural analysis of the cDNA and the gene encoding the murine type 1 sigma receptor. Biochem Biophys Res Commun 1997;241:535–40.
- Seth P, Fie YJ, Li HW, Huang W, Leibach FH, Ganapathy V. Cloning and functional characterization of a sigma receptor from rat brain. J Neurochem 1998;70:922–31.
- Sherman JE, Jorenby DE, Baker TB. Classical conditioning with alcohol: acquired preferences and aversions, tolerance and urge/craving. In: Chaudron CD, Wilkinson DA, editors. Theories on alcoholism. Toronto: Addiction Research Foundation; 1998. p. 173–237.
- Su TP, Hayashi T. Cocaine affects the dynamics of cytoskeletal proteins via sigma(1) receptors. Trends Pharmacol Sci 2001;22:456-8.
- Su TP, Wu XZ, Cone EJ, Shukla K, Gund TM, Dodge AL, et al. Sigma compounds derived from phencyclidine: identification of PRE-084, a new, selective sigma ligand. J Pharmacol Exp Ther 1991;259:543–8.
- Tabakoff B, Hoffman PL. Alcohol addiction: an enigma among us. Neuron 1996;16:909-12.
- Tzschentke TM. Measuring reward with the conditioned place preference paradigm: a comprehensive review of drug effects, recent progress and new issues. Prog Neurobiol 1998;56:613–72.
- Ujike H, Kanzaki A, Okumura K, Akiyama K, Otsuka S. Sigma (σ) antagonist BMY 14802 prevents methamphetamine-induced sensitization. Life Sci 1992;50:PL129–34.
- Ujike H, Kuroda S, Otsuka S. σ Receptor antagonists block the development of sensitization to cocaine. Eur J Pharmacol 1996;296:123-8.
- Weiss F, Lorang MT, Bloom FE, Koob GF. Oral alcohol self-administration stimulates dopamine release in the rat nucleus accumbens: genetic and motivational determinants. J Pharmacol Exp Ther 1993;267:250–8.
- Wise RA, Bozarth MA. A psychomotor stimulant theory of addiction. Psychol Rev 1987;94:469–92.
- Witkin JM, Terry M, Menkel M, Hickey P, Pontecorvo M, Ferkany J, et al. Effects of the selective sigma receptor ligand 6-[6-(4-hydroxypiperidinyl)hexyloxy]-3-methylflavone (NPC 16377), on behavioral and toxic effects of cocaine. J Pharmacol Exp Ther 1993;266:473–82.
- Zamanillo D, Andreu F, Ovalle S, Perez MP, Romero G, Farre AJ, et al. Up-regulation of sigma<sub>1</sub> receptor mRNA in rat brain by a putative atypical antipsychotic and sigma receptor ligand. Neurosci Lett 2000; 282:169–72.